ASIC-dependent LTP at multiple glutamatergic synapses in amygdala network is required for fear memory

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Supplemental Information

Supplemental Figures



Supplementary Figure S1 | Classification of GABAergic INs in the BLA

(**a-d**), Histograms of accommodating ratios, maximal CVs of inter-spike interval (ISI) ratios, delays of AP and maximal mean firing rates from GFP⁺ cells in the BLA. The D'Agostino and Pearson omnibus normality test demonstrates that the properties presented are not unimodally distributed. Asterisks indicate significant deviation from a normal distribution (*P < 0.05, **P < 0.01, ***P < 0.001). (**e**) Hierarchical cluster analysis of GABAergic INs performed with electrophysiological parameters shown in **a-d** as the parameters for classification. The x axis of the dendrogram represents the individual cells, and the y axis represents the rescaled distance (squared Euclidean) between groups. Distinct subtypes of INs are marked with bars below the dendrogram. Inset, pie chart shows proportion of each BLA-IN subtype. (**f-i**), Bar graphs comparing accommodating ratios, maximal CVs of ISI ratios, delays of AP and maximal firing rates among different IN subtypes ($^{*}P < 0.05$, $^{***}P < 0.001$, Wilcoxon rank-sum test after Kruskal-Wallis test).



Supplementary Figure S2 | Classification of GABAergic neurons in the CeL

(**a**,**b**), Membrane responses to supra- (black) and sub-threshold (dark and light green) current pulse injections recorded from a representative LS (**a**) and ES cell (**b**). Note the slow depolarizing ramp of the membrane response in the LS cell but not in the ES cell. Bars a and b indicate how the average membrane potentials were measured for the calculation of ramp ratio (b/a). (**c**-**f**), Histograms of spike delays, ramp ratios, rheobases and resting membrane potentials (RMPs) from non-LTB cells. Asterisks indicate significant deviation from a normal distribution (*P < 0.05, **P < 0.01, D'Agostino and Pearson omnibus normality test). (g) Dendrogram of non-LTB neurons. The y axis represents the normalized Euclidean distance. Color codes indicate cell classification based on spike delay; cells with spike delay > 1.5 s were dark green otherwise light green. Inset, pie chart shows distinct populations in the CeL. Dark and light green areas denote the major cell populations, LS and ES neurons. Gray area depicts a small subset of LTB neurons. (h) Scatter-plot of spike delay vs. ramp ratio. The dotted line indicates the arbitrary cutoff value (spike delay of 1.5 s) that best separates the two populations.



Supplementary Figure S3 | ASIC current kinetics of different amygdala neurons

(a) Top, a representative ASIC current trace (black; average of 8 trials) from a BLA-PN; red curve is an exponential function fit. Bottom, bar graph showing desensitization τ of each cell type. P = 0.52, Kruskal-Wallis test. Numbers of patches are given in parentheses above bars. (b) Heat map summarizing the desensitization τ s of ASIC currents of different amygdala neurons. (c) Top, representative traces showing the recovery ratio was measured at -65 mV after rapid changes from pH 7.4 to 5 for 1 s and then changed back to pH 7.4 for 20 s, followed by a second pulse to pH 5 for 1 s. Bottom, bar graph showing the recovery (I_2/I_1) ratio of ASIC currents of each cell type. P = 0.65, Kruskal-Wallis test. Numbers of patches are given in parentheses above bars. (d) Heat map summarizing the recovery ratios of ASIC currents of different amygdala neurons. (e-f) Representative traces showing the recovery from desensitization, which were recorded from BLA-PNs (e) and CeM-LTB neurons (f), was measured with 5 s (left) and 20 s (right) inter-pulse intervals. (g) Data points obtained from BLA-PN and CeM-LTB were fitted with a monoexponential function, yielding recovery time constants of 10 s and 18 s, respectively.

			BLA			
	I PN (23)	II AcIN (15)	III StIN (11)	IV DFIN (10)	V FSIN (10)	P value
RMP (mV)	-67.1 ± 1.3	-60.1 ± 1.5	-59.6 ± 2.7	-65.5 ± 2.0	$\textbf{-68.0} \pm 2.4$	< 0.001
	II, III, IV	I, IV, V	I, IV, V	I, II, III	II, III	
R_{in} (M Ω)	269 ± 40	330 ± 41	298 ± 57	244 ± 28	242 ± 38	0.16
Sag ratio (%)	22.9 ± 2.3	25.3 ± 4.3	24.3 ± 3.6	15.1 ± 2.3	22.8 ± 6.2	0.23
Rheobase (pA)	72.1 ± 9.7	59.5 ± 11.2	81.9 ± 24.9	91.7 ± 16.0	73.0 ± 19.2	0.45
Accommodating	1.89 ± 0.01	3.44 ± 0.23	2.79 ± 0.33	1.65 ± 0.12	1.43 ± 0.06	< 0.001
ratio	II, V	I, IV, V	IV, V	II, III	I, II, III	
Maximal CV of	0.42 ± 0.07	0.37 ± 0.03	1.71 ± 0.11	0.41 ± 0.10	0.13 ± 0.04	< 0.001
ISI ratios	III, V	III, V	I, II, IV, V	III, V	I, II, III, IV	
Spike delay	237 ± 54	121 ± 16	134 ± 23	677 ± 48	133 ± 41	< 0.001
(ms)	IV	IV	IV	IV	IV	
Maximal mean	28.3 ± 1.3	33.8 ± 2.9	45.2 ± 6.2	38.6 ± 3.7	100.6 ± 9.6	< 0.001
firing rate (Hz)	II, III, IV, V	V	V	V	V	
AP threshold	-37.1 ± 1.4	-39.3 ± 2.3	-38.4 ± 1.7	-34.0 ± 1.5	-42.3 ± 1.4	0.011
(mV)	IV	IV	IV	I, II, III, V	IV	
AP half-width	1.54 ± 0.11	0.82 ± 0.05	0.78 ± 0.05	1.18 ± 0.07	0.64 ± 0.07	< 0.001
(ms)	II, III, IV, V	I, IV	I, IV	I, II, III, V	I, IV	
AP amplitude	98.6 ± 1.3	91.7 ± 2.5	82.3 ± 2.6	74.6 ± 1.5	90.6 ± 2.9	< 0.001
(mV)	II, III, IV, V	I, III, IV	I, II, IV	I, II, III, V	I, II, IV	
AP maximum	320 ± 20	311 ± 17	270 ± 18	210 ± 16	327 ± 22	0.001
rise (V/s)	IV	IV	IV	I, II, III, V	IV	
AP maximum	-57 ± 15	-105 ± 13	-111 ± 10	-59 ± 11	-184 ± 8	< 0.001
decay (V/s)	II, III, V	I, IV, V	I, IV, V	II, III, V	I, II, III, IV	

Supplemental Tables Supplementary Table S1 | Electrophysiological properties of neurons in the BLA

Numbers of cells are given in parentheses. Kruskal-Wallis test was performed to compare the means among groups. All values are given as mean \pm s.e.m. Statistical differences between PNs and four types of INs, estimated based on Wilcoxon rank-sum test after Kruskal-Wallis test, are marked under the corresponding parameter, P < 0.05. RMP, resting membrane potential; R_{in}, input resistance.

	C		
	LS (46)	ES (26)	P value
RMP (mV)	-70.8 ± 1.4	-65.7 ± 2.2	0.033
\mathbf{R}_{in} (M Ω)	420 ± 25	415 ± 36	0.038
Sag ratio (%)	17.6 ± 2.3	24.3 ± 3.1	0.017
Rheobase (pA)	37.6 ± 2.7	31.8 ± 3.0	0.17
Spike delay (ms)	1769 ± 24	778 ± 86	< 0.001
Ramp ratio	1.37 ± 0.02	0.93 ± 0.02	< 0.001
AP threshold (mV)	-34.8 ± 0.7	-33.1 ± 1.1	0.30
AP half-width (ms)	2.03 ± 0.11	1.63 ± 0.13	< 0.001
AP amplitude (mV)	89.3 ± 1.1	90.5 ± 2.3	0.11
AP maximum rise (V/s)	218 ± 9	241 ± 14	0.06
AP maximum decay (V/s)	-42 ± 44	-54 ± 25	0.008

Supplementary Table S2 | Electrophysiological properties of the major cell types in the CeL

Wilcoxon rank-sum test was performed to determine statistical significance between the groups.

Numbers of cells are given in parentheses. All values are given as mean \pm s.e.m.

	CeM		
	LTB (37)	ES (29)	P value
RMP (mV)	-58.0 ± 1.2	-61.7 ± 2.0	0.07
R_{in} (M Ω)	568 ± 42	589 ± 50	0.71
Sag ratio (%)	43.7 ± 4.0	18.0 ± 1.8	< 0.001
Rheobase (pA)	21.1 ± 1.8	33.2 ± 4.0	0.013
AP threshold (mV)	-39.5 ± 0.9	-35.8 ± 1.2	0.007
AP half-width (ms)	1.17 ± 0.03	1.33 ± 0.07	0.06
AP amplitude (mV)	92.5 ± 1.2	91.1 ± 1.6	0.52
AP maximum rise (V/s)	254 ± 9	248 ± 10	0.59
AP maximum decay (V/s)	-74 ± 38	-66 ± 29	0.05

Supplementary Table S3 | Electrophysiological properties of the major cell types in the CeM

Wilcoxon rank-sum test was performed to determine the statistical significance between the groups.

Numbers of cells are given in parentheses. All values are given as mean \pm s.e.m.

	ICM _{MV} (16)	ICM _{MD} (12)	ICM _L (8)	P value
RMP (mV)	-65.3 ± 2.2	-66.3 ± 1.3	-71.1 ± 2.5	0.10
$R_{in}(M\Omega)$	693 ± 77	709 ± 60	692 ± 72	0.73
Sag ratio (%)	18.1 ± 2.3	18.5 ± 4.6	21.6 ± 5.0	0.54
Rheobase (pA)	32.6 ± 3.4	34.9 ± 5.9	37.3 ± 3.3	0.42
AP threshold (mV)	-32.9 ± 1.4	-33.5 ± 1.2	-31.8 ± 2.6	0.90
AP half-width (ms)	1.14 ± 0.06	1.08 ± 0.04	1.21 ± 0.05	0.25
AP amplitude (mV)	86.9 ± 1.9	88.4 ± 1.9	85.6 ± 2.7	0.48
AP maximum rise (V/s)	228 ± 12	227 ± 10	200 ± 15	0.28
AP maximum decay (V/s)	-72 ± 16	-76 ± 13	-66 ± 9	0.10

Supplementary Table S4 | Electrophysiological properties of neurons in the ICMs

Numbers of cells are given in parentheses. Kruskal-Wallis test was performed to compare the

means among groups. All values are given as mean \pm s.e.m.